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Asymmetric synthesis of β -amino- α -hydroxyphosphinic acid derivatives through hydrophosphinylation of α -amino aldehydes

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Abstract—The diastereoselective synthesis of β-amino-α-hydroxyphosphinates was achieved by hydrophosphinylation of N,N-dibenzyl-α-amino aldehydes with ethyl ethylphosphinate catalyzed by (S)-ALB. The hydrophosphinylation using ethyl phosphinate afforded both syn-and anti-β-amino-α-hydroxy-H-phosphinates with high diastereoselectivities by tuning the chirality of ALB. © 2002 Elsevier Science Ltd. All rights reserved.

The β -amino- α -hydroxyphosphinic acids **1** serve as the key intermediates for the synthesis of potent inhibitors of human renin and HIV protease (Fig. 1). Let 2. Stereogenic carbon-phosphorus bond formation processes are of great interest in the stereoselective synthetic sequences of the β -amino alcohol moiety by the reaction of α -amino aldehydes with phosphinic nucleophiles, since the stereochemistry of β -amino alcohol is known to be an important factor to get potent inhibitory active compounds. Although β -amino- α -hydroxyphosphinic acid derivatives were obtained by the reaction of *N*-Boc- α -amino aldehydes with methyl ethylphosphinate (HPO(OMe)(Et)) in the presence of TMSCI and an amine, diastereoselectivity was not observed. Our special interest is in developing concise and highly diastereoselective synthesis of both β -amino- α -hydroxy-

$$H_2N$$
 H_2N
 H_2N

Figure 1.

phosphinic acids (1/X=H). We examined the chiral AlLibis(binaphthoxide) (ALB)³-catalyzed hydrophosphinylayion of α-amino aldehydes employing two types of phosphinic nucleophile, alkyl alkylphosphinate (HPO(OR)(R)) and alkyl phosphinate (H₂PO₂R), in extension of our previous work on catalytic asymmetric hydrophosphinylation of aldehydes.⁴ The reaction of ethyl ethylphosphinate⁵ with N,N-dibenzyl- α -amino aldehydes⁶ afforded anti-β-amino-α-hydroxyphosphinates in a highly diastereoselective manner. Furthermore, chiral ALBcatalyzed hydrophosphinylation employing ethyl phosphinate, generated from anhydrous phosphinic acid and triethyl orthoformate in situ, proceeded with a high level of diastereofacial selectivity. The stereochemical outcome of the reaction can be controlled in either anti- or synselective manner by tuning the chirality of ALB.8 We wish to describe full details in this paper (Scheme 1).

phosphinic acids (1/X=alkyl) and β-amino-α-hydroxy-H-

First, we examined the hydrophosphinylation of N,N-dibenzyl- α -amino aldehyde **2a** with ethyl ethylphosphinate in the presence of (R)-ALB (20 mol%), generated from (R)-binaphthol, in THF at -40° C for 12 h (Table 1). The reaction was found to be sluggish, giving adducts syn-**3** and anti-**3** only in 4% yield (entry 1). The ³¹P NMR spectrum of

Scheme 1.

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Table 1. Hydrophosphinylation of 2a with ethyl ethylphosphinate in the presence of ALB

Entry ^a	ALB	Temperature (°C)	syn/anti ^b	Yield (%) ^c	
1	(R)-ALB	-40	43:57	4	
2	(R)-ALB	0	43:57	55	
3	(S)-ALB	0	11:89	51	

^a All reactions were carried out for 12 h.

the crude products revealed that the diastereoselectivity was poor (syn/anti=43:57). When the reaction was carried out at 0°C, the chemical yield was raised to 55%, but resulting in the same diastereoselectivity (entry 2). On the other hand, employing (S)-ALB gave adduct 3 with anti-selectivity (syn/anti=11:89) in 51% yield (entry 3). These results indicated that the combination of (S)-ALB and 2a was suitably matched for inducing diastereofacial selectivity. The hydrophosphinylation products syn-3 and anti-3 were obtained as a 1:1 mixture of isomers arising from the chirality of the phosphinate group, respectively. Only in the case of syn-3, these diastereomers, syn-3-A and syn-3-B, were separated by silica gel column chromatography.

In view of the level of selectivity, ALB-catalyzed hydrophosphinylation was confirmed to be more advantageous over the existing conventional method; that is, hydrophosphinylation of **2a** using 1.5 equiv. of both Et₃N and TMSCl, followed by desilylation with Bu₄NF afforded **3** with modest selectivity (*syn/anti*=30:70) in 55% yield (Scheme 2). The stereochemistry of the major diastereomer

Scheme 2.

was estimated to be *anti* by considering the Felkin–Anh transition state. ¹⁰

We next examined hydrophosphinylation of 2a,b employing ethyl phosphinate for the synthesis of β -amino- α -hydroxy-H-phosphinates in a highly diastereoselective manner (Table 2). In these cases, ethyl phosphinate is expected to be readily activated by ALB showing increased nucleophilicity due to the low pK_a value in comparison with ethyl ethylphosphinate.

As we expected, ethyl phosphinate could be activated by the catalyst even at -40°C affording the corresponding hydrophosphinylation products in moderate yields. 11 An intriguing result was revealed upon analyzing the diastereoselectivities of the products. Performing the reaction of 2a in the presence of (R)-ALB gave rise to high syn-selectivity (syn-4a/anti-4a=87:13) (entry 1). On the other hand, the reaction of 2a by the use of (S)-ALB instead of (R)-ALB proceeded with inversed stereoselection to afford anti-4a in a ratio of 6:94 (entry 2). Also, when the reaction of **2b** was carried out with (R)- or (S)-ALB, either syn-**4b** or anti-**4b** could be obtained in high stereoselectivity (entries 3 and 4). In the above-mentioned cases, diastereofacial selectivity was found to be controlled predominantly by the chirality of the asymmetric catalyst rather than that of the α -amino aldehydes. As observed in Table 2, the diastereoselectivity

Table 2. Hydrophosphinylation of 2a,b with ethyl phosphinate in the presence of ALB

$$\begin{array}{c} NBn_2 \\ \hline P \\ CHO \end{array} \begin{array}{c} O \\ H-P-H \\ \hline OEt \\ \hline (S)- \text{ or } (R)\text{-ALB} \\ (20 \text{ mol }\%) \\ \hline THF, -40 °C \end{array} \begin{array}{c} Bn_2 N \\ \hline P-H \\ \hline HO \\ \hline OEt \\ \hline HO \\ \hline Syn-4a,b \end{array} \begin{array}{c} Bn_2 N \\ \hline P-H \\ \hline HO \\ \hline Anti-4a,b \\ \hline \end{array}$$

a: R = CH₂Ph; b: R = i-Bu

Entry ^a	Substrate	ALB	syn/anti	Yield (%) ^b
1	2a	(R)-ALB	87:13	66
2	2a	(S)-ALB	6:94	56
3	2b	(R)-ALB	94:6	54
4	2b	(S)-ALB	2:98	71

^a All reactions were carried out for 12 h at -40° C.

^b Determined by ³¹P NMR analysis of crude products.

^c Combined yields of syn- and anti-isomers.

^b Combined yields of syn- and anti-isomers.

Figure 2.

Scheme 3.

of the hydrophosphinylation with (*S*)-ALB is generally higher than that with (*R*)-ALB.

The high diastereoselectivity of the hydrophosphinylation catalyzed by ALB would be accounted for by the kinetically controlled process: treatment of anti-4a with ethyl phosphinate in the presence of (R)-ALB at -40° C formed none of syn-isomer that would be expected from a reversible process. Although the exact reason for high selectivities in (R)-ALB-catalyzed hydrophosphinylation using ethyl phosphinate in comparison with ethyl ethylphosphinate remains unclear, it seems likely to be associated with a steric disposition for phosphinic nucleophile.

The products *anti*-**4a,b** and *syn*-**4a,b** were obtained as a 1:1 mixture of diastereoisomers arising from the chirality of the phosphinate group. The diastereomerically pure *anti*-**4a-A** (mp: 149–150°C) was isolated from the mixture (*anti*-**4a-A** and *anti*-**4a-B**) upon recrystallization from ethyl acetate. The relative stereochemistry of *anti*-**4a-A** was confirmed unambiguously by X-ray crystallographic analysis (Fig. 2).

The stereochemistry of *anti*-4a,b was also confirmed after converting to β -amino- α -acetoxyphosphonate 6a,b through sequential acetylation, oxidation, ¹³ deesterification ¹⁴ and methyl esterification (Scheme 3). The ¹H NMR spectra of 6a,b were identical with those of the authentic specimens derived from the known β -amino- α -hydroxyphosphonate 7a,b^{10,15} through acetylation and deesterification followed by methyl esterification. The optical purity of 6a derived from *anti*-4a was determined to be 99% ee by HPLC

analysis on a chiral phase (DAICEL CHIRALPAK OD column, hexane/EtOH=20:1). Therefore, it was proved that no racemization of N,N-dibenzyl- α -amino aldehydes took place during the hydrophosphinylation.

Finally, the ethyl ester of *anti-3* could be easily removed by treatment with TMSBr followed by methanolysis to give β-amino-α-hydroxyphosphinic acid **9** in 92% yield (Scheme 4) The product **9** was obtained as a single product due to the loss of asymmetric character of phosphorus atom by the rapid transfer of the acidic proton between the phosphoryl (P=O) and the acidic (P-OH) sites. ¹³

In conclusion, we have developed a diastereoselective synthesis of β -amino- α -hydroxyphosphinates through hydrophosphinylation of N,N-dibenzyl- α -amino aldehydes with ethyl ethylphosphinate catalyzed by (S)-ALB. Moreover, applying ethyl phosphinate to the hydrophosphinylation afforded both syn- and anti- β -amino- α -hydroxy-H-phosphinates selectively by tuning the chirality of ALB. The present methodology would be widely applicable to the synthesis of protease inhibitors.

1. Experimental

All melting points were taken on a Yanagimoto micromelting point apparatus and are uncorrected. Optical rotations were measured with a JASCO DIP-360 digital polarimeter. IR spectra were recorded on a JASCO FTIR-620. Mass spectra were measured on a Finnigan TSQ-700 or a VG Auto Spec E. Elemental analysis were recorded on an Elemental Vavio EL. NMR spectra were obtained on either a Bruker DPX400 NMR spectrometer operating or a Varian Mercury-300BB instrument operating at 400 (or 300) MHz for 1 H, 100 (or 75.5) MHz for 13 C, and 162 MHz for 31 P. The chemical shift data for each signal on 1 H NMR are given in units of δ relative to CHCl₃ (δ =7.26) for CDCl₃

solution or CH₃OH (δ =3.30) for CD₃OD solution. For ¹³C NMR spectra, the chemical shifts in CDCl₃ or CD₃OD are reported relative to the CDCl₃ resonance (δ =77.0) and CD₃OD resonance (δ =49.0), respectively. The chemical shifts of ³¹P are recorded relative to external 85% H₃PO₄ (δ =0) with broad-band ¹H decoupling. The aldehydes **2a,b** were prepared in enantiomerically pure forms from the corresponding L-amino acids according to the literature methods and used without purification. ⁶ All reactions were conducted under nitrogen.

1.1. The procedure for the hydrophosphinylation of 2a with ethyl ethylphosphinate in the presence of (R)-ALB

To a solution of ethyl ethylphosphinate (732 mg, 6.0 mmol) in THF (4 mL) was added 0.1 M THF solution of (*R*)-ALB (8.0 mL, 0.8 mmol), prepared from (*R*)-BINOL (458 mg, 1.6 mmol) and LiAlH₄ (30.4 mg, 0.8 mmol) in situ according to Shibasaki's method,³ and a solution of **2a** (4.0 mmol) in THF (8 mL) at 0°C under stirring. After stirring for 12 h at the same temperature, the mixture was diluted with H₂O and extracted with CHCl₃. The combined extracts were washed with brine and dried over MgSO₄. Removal of the solvent gave a residue which was chromatographed on silica gel (hexane/EtOAc=1:1 to CHCl₃/MeOH=20:1) to give *syn*-**3-A** (142 mg, 8%), *syn*-**3-B** (338 mg, 19%) and *anti*-**3** (513 mg, 28%).

- **1.1.1.** Ethyl (1*S*,2*S*)-2-(dibenzylamino)-1-hydroxy-3-phenylpropyl(ethyl)phosphinate (*syn*-3-A). Oil; $[\alpha]_D^{23}$ = +24.2 (*c* 0.84, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.42–7.04 (15H, m), 5.09 (1H, d, J=15.8 Hz), 4.19–4.06 (2H, m), 3.77 (2H, d, J=12.6 Hz), 3.66 (1H, dd, J=5.4, 10.7 Hz), 3.50–3.33 (3H, m), 2.98 (1H, dd, J=10.5, 14.4 Hz), 1.55–1.14 (2H, m), 1.35 (3H, t, J=7.0 Hz), 0.87 (3H, td, J=7.8, 17.3 Hz); ³¹P NMR (162 MHz, CDCl₃) δ 58.76; ¹³C NMR (75.5 MHz, CDCl₃) δ 140.0, 138.0, 129.6, 129.3, 128.6, 128.5, 127.4, 126.5, 66.4 (d, J_{PC}=110.6 Hz), 60.3 (d, J_{PC}=6.3 Hz), 58.6, 58.5, 53.3, 34.9, 16.5 (d, J_{PC}=93.9 Hz), 16.3 (d, J_{PC}=5.2 Hz), 5.4 (d, J_{PC}=5.8 Hz); IR (neat) 3240, 1029 cm⁻¹; EIMS m/z 360 (M⁺ CH₂Ph). High resolution MS calcd for C₂₀H₂₇NO₃P (M⁺ CH₂Ph): 360.1728. Found: 360.1722.
- 1.1.2. Ethyl (1S,2S)-2-(dibenzylamino)-1-hydroxy-3-phenylpropyl(ethyl)phosphinate (syn-3-B). Mp $^{102-104^{\circ}\text{C}}$; 1 [α] $_{\text{D}}^{^{24}}$ = +35.9 (c 0.93, CHCl₃); 1 H NMR (300 MHz, CDCl₃) δ 7.41–7.04 (15H, m), 4.94 (1H, d, J=17.0 Hz), 3.82–3.66 (4H, m), 3.55–3.45 (3H, m), 3.34–3.30 (1H, m), 3.01 (1H, dd, J=10.7, 14.8 Hz), 1.93–1.71 (2H, m), 1.17 (3H, td, J=7.8, 17.9 Hz), 1.03 (3H, t, J=7.0 Hz); 31 P NMR (162 MHz, CDCl₃) δ 55.80; 13 C NMR (75.5 MHz, CDCl₃) δ 140.2, 138.1, 129.6, 129.3, 128.6, 128.4, 127.4, 126.5, 66.6 (d, J_{PC}=111.7 Hz), 61.0 (d, J_{PC}=7.5 Hz), 58.4, 53.3 (2 carbons), 35.0, 19.4 (d, J_{PC}=91.6 Hz), 16.6 (d, J_{PC}=5.8 Hz), 5.3 (d, J_{PC}=5.2 Hz); IR (KBr) 3167, 1029 cm $^{-1}$; EIMS m/z 452 (MH $^{+}$). Anal. Calcd for C₂₇H₃₄NO₃P: C, 71.82; H, 7.59. Found: C, 71.68; H, 7.59.
- **1.1.3.** Ethyl (1*R*,2*S*)-2-(dibenzylamino)-1-hydroxy-3-phenylpropyl(ethyl)phosphinate (*anti*-3). This compound was obtained as a mixture of diastereomers in a ratio of 1:1.

Mp 110–113°C; ¹H NMR (400 MHz, CDCl₃) δ 7.30–7.06 (15H, m), 4.26–4.10 (1H, m), 4.09–3.98 (2H, m), 3.89 (1H, d, J=14.1 Hz), 3.83 (1H, d, J=14.1 Hz), 3.70 (1H, d, J=14.1 Hz), 3.58 (1H, d, J=14.1 Hz), 3.40–3.36 (1H, m), 3.12–3.07 (2H, m), 1.84–1.59 (2H, m), 1.23 (1.5H, t, J=7.0 Hz), 1.19 (1.5H, t, J=7.0 Hz), 1.16 (1.5H, td, J=7.7, 17.4 Hz), 1.06 (1.5H, td, J=7.8, 17.4 Hz); ³¹P NMR (162 MHz, CDCl₃) δ 54.61, 54.39; IR (KBr) 3261, 1045 cm⁻¹; EIMS m/z 360 (M⁺ – CH₂Ph). Anal. Calcd for C₂₇H₃₄NO₃P: C, 71.82; H, 7.59. Found: C, 71.41; H, 7.57.

1.2. The procedure for the hydrophosphinylation of 2a with ethyl ethylphosphinate in the presence of TMSCl and Et_3N

To a solution of ethyl ethylphosphinate (732 mg, 6.0 mmol) in CH₂Cl₂ (21 mL) was added TMSCl (0.76 mL, 6.0 mmol) and Et₃N (0.83 mL, 6.0 mmol) at 0°C and the mixture was stirred for 0.5 h at the same temperature. To this solution was added a solution of 2a (4 mmol) in THF (4 mL) at 0°C and the mixture was stirred for 12 h at the same temperature. The mixture was poured into cold H₂O and extracted with CHCl₃. The combined extracts were washed with brine and dried over MgSO₄. Removal of the solvent gave a residue. To a solution of the residue in THF (8 mL) was added 1 M THF solution of Bu₄NF (4.8 mL, 4.8 mmol) at 0°C. After stirring for 1 h at the same temperature, the mixture was poured into cold H₂O and extracted with CHCl₃. The combined extracts were washed with brine and dried over MgSO₄. Removal of the solvent gave a residue which was chromatographed on silica gel (hexane/EtOAc=1:1 to CHCl₃/MeOH=20:1) to give a mixture of syn-3-A, syn-3-**B** and *anti-***3** (992 mg, 55%).

1.3. General procedure for the hydrophosphinylation of 2a,b with ethyl phosphinate in the presence of (R)-ALB

A mixture of anhydrous phosphinic acid (396 mg, 6.0 mmol) and triethyl orthoformate (3.0 mL, 18.0 mmol) was stirred for 1.5 h at room temperature. To this solution was added 4 mL of THF and 0.1 M THF solution of (*R*)-ALB (8.0 mL, 0.8 mmol), prepared from (*R*)-BINOL (458 mg, 1.6 mmol) and LiAlH₄ (30.4 mg, 0.8 mmol) in situ, and a solution of **2a,b** (4.0 mmol) in THF (8 mL) at -40°C under stirring. After stirring for 12 h at the same temperature, the mixture was diluted with H₂O and extracted with CHCl₃. The combined extracts were washed with brine and dried over MgSO₄. Removal of the solvent gave a residue which was chromatographed on silica gel (hexane/EtOAc=1:1 to AcOEt) to give *syn-4a,b* and *anti-4a,b*.

1.3.1. Ethyl (1S,2S)-(dibenzylamino)-1-hydroxy-3-phenyl-propylphosphinate (*syn-4a*). This compound was obtained as a mixture of diastereomers in a ratio of 1:1. One of the diastereisomers is crystallized in another oily diastereisomer. Yield (987 mg, 58%); 1 H NMR (400 MHz, CD₃OD) δ 7.44–7.16 (15H, m), 7.15 (0.5H, d, J=561.7 Hz), 7.03 (0.5H, d, J=560.3 Hz), 4.28–4.18 (2H, m), 4.14–4.04 (2H, m), 3.89 (0.5H, m), 3.77–3.54 (3H, m), 3.42–3.36 (0.5H, m), 3.26–3.07 (1.5H, m), 2.88–2.83 (0.5H, m), 1.30 (3H, t, J=7.0 Hz); 31 P NMR (162 MHz, CD₃OD) δ 44.42, 40.09; IR (KBr) 3221, 1190 cm⁻¹;

FABMS m/z 424 (MH⁺). Anal. Calcd for C₂₅H₃₀NO₃P: C, 70.90; H, 7.14. Found: C, 71.08; H, 7.10.

- Ethyl (1R,2S)-(dibenzylamino)-1-hydroxy-3phenylpropylphosphinate (anti-4a). This compound was obtained as a mixture of diastereomers in a ratio of 1:1. One of the diastereisomers is crystallized in another oily diastereisomer. Yield (131 mg, 8%); ¹H NMR (400 MHz, CD₃OD) δ 7.27–7.08 (15H, m), 6.68 (0.5H, d, J=549.5 Hz), 6.53 (0.5H, d, J=558.0 Hz), 4.42–4.34 (1H, m), 4.28–3.95 (2H, m), 3.91 (1H, d, *J*=12.3 Hz), 3.86 (1H, d, J=14.0 Hz), 3.78-3.70 (0.5 H, m), 3.60 (1 H, d, d)J=14.3 Hz), 3.52 (1H, d, J=14.1 Hz), 3.49-3.44 (0.5H, m), 3.20-3.11 (1.5H, m), 2.99 (0.5H, dd, *J*=4.6, 14.3 Hz), 1.28 (1.5H, t, J=7.1 Hz), 1.18 (1.5H, t, J=7.0 Hz); ³¹P NMR (162 MHz, CD₃OD) δ 40.66, 38.40; IR (KBr) 3267, 1197 cm $^{-1}$; FABMS m/z 424 (MH $^{+}$). Anal. Calcd for C₂₅H₃₀NO₃P: C, 70.90; H, 7.14. Found: C, 70.83; H, 7.12.
- **1.3.3.** Ethyl (1*S*,2*S*)-(dibenzylamino)-1-hydroxy-4-methylpentylphosphinate (*syn*-4b). This compound was obtained as a mixture of diastereomers in a ratio of 1:1. Yield (788 mg, 51%); An oil; ¹H NMR (400 MHz, CD₃OD) δ 7.39–7.24 (10H, m), 7.12 (0.5H, d, J=558.6 Hz), 7.03 (0.5H, d, J=555.5 Hz), 4.27–4.17 (1H, m), 4.15–4.01 (2H, m), 3.79–3.53 (4H, m), 3.31–3.25 (0.5H, m), 3.17–3.12 (0.5H, m), 1.80–1.60 (2H, m), 1.39 (1.5H, t, J=7.1 Hz), 1.29 (1.5H, t, J=7.0 Hz), 1.23–1.19 (1H, m), 0.93 (1.5H, t, J=6.6 Hz), 0.85 (1.5H, d, J=6.5 Hz), 0.82 (1.5H, d, J=6.1 Hz), 0.79 (1.5H, d, J=6.5 Hz); ³¹P NMR (162 MHz, CD₃OD) δ 43.64, 41.92; IR (neat) 3255, 1189 cm⁻¹; EIMS m/z 296 (M⁺−HP(O)OEt). High resolution MS calcd for C₂₀H₂₆NO (M⁺−HP(O)OEt): 296.2014. Found: 296.2002.
- **1.3.4.** Ethyl (1*R*,2*S*)-(dibenzylamino)-1-hydroxy-4-methylpentylphosphinate (*anti*-4b). This compound was obtained as a mixture of diastereomer in a ratio of 1:1. Yield (44 mg, 3%); An oil; 1 H NMR (400 MHz, CD₃OD) δ 7.38–7.24 (10H, m), 6.97 (0.5H, d with small splits, J=540.3 Hz), 6.94 (0.5H, d, J=542.3 Hz), 4.45–4.38 (1H, m), 4.24–4.10 (2H, m), 3.95 (2H, d, J=13.6 Hz), 3.47 (2H, d, J=13.6 Hz), 3.22–3.16 (1H, m), 1.97–1.88 (2H, m), 1.39 (1.5H, t, J=7.0 Hz), 1.33 (1.5H, t, J=7.0 Hz), 1.29–1.24 (1H, m), 0.94 (3H, d, J=6.7 Hz), 0.53 (1.5H, d, J=6.2 Hz), 0.51 (1.5H, d, J=6.3 Hz); 31 P NMR (162 MHz, CD₃OD) δ 40.32, 39.00; IR (neat) 3277, 1203 cm⁻¹; FABMS m/z 390 (MH⁺). High resolution MS calcd for C₂₂H₃₂NO₃P (MH⁺): 390.2198. Found: 390.2175.
- **1.3.5. Ethyl (1***R*,**2***S*)-(dibenzylamino)-1-hydroxy-3-phenyl-propylphosphinate (*anti*-4a-A). This compound was obtained by recrystallization of *anti*-4a from EtOAc. Mp 149–150°C; $[\alpha]_D^{25}$ =+13.7 (*c* 0.80, MeOH); ¹H NMR (400 MHz, CD₃OD) δ 7.17–7.08 (15H, m), 6.68 (1H, d, *J*=549.5 Hz), 4.38–4.32 (1H, m), 4.11–3.98 (2H, m), 3.86 (2H, d, *J*=14.0 Hz), 3.52 (2H, d, *J*=14.1 Hz), 3.49–3.44 (1H, m), 3.12 (1H, dd, *J*=9.7, 14.3 Hz), 2.99 (1H, dd, *J*=4.6, 14.3 Hz), 1.28 (3H, t, *J*=7.1 Hz); ³¹P NMR (162 MHz, CD₃OD) δ 40.66; ¹³C NMR (100 MHz, CD₃OD) δ 141.4, 140.8, 131.0, 130.0, 129.1, 127.9, 127.1, 68.6 (d, *J*_{PC}=109.3 Hz), 64.5 (d, *J*_{PC}=7.9 Hz), 60.3 (d, *J*_{PC}=8.0 Hz), 55.3 (2 carbons), 34.4, 16.7 (d,

 J_{PC} =5.9 Hz); IR (KBr) 3271, 1197 cm⁻¹; FABMS m/z 424 (MH⁺). Anal. Calcd for $C_{25}H_{30}NO_3P$: C, 70.90; H, 7.14. Found: C, 70.91; H, 7.17.

1.4. Crystal data for compound anti-4a-A

X-Ray crystal data of *anti*-**4a-A** were collected by Mac-Science MXC18 diffractometer. The structure was solved by a direct method using SIR92¹⁶ and refined with a full matrix least-squares method. Molecular formula= $C_{25}H_{30}$. NO₃P, M_r =423.50, Orthorhombic, space group= $P2_12_12_1$, a=16.725 (4) Å, b=12.170 (2) Å, c=11.434 (3) Å, V=2327.2 (9) Å³, T=298 K, Z=4, D_x =1.208 mg m⁻³, (Mo K α)=0.71073 Å, μ =1.375 mm⁻¹, R=0.083 over 2658 independent reflections. Crystallographic data (excluding structure factors) for the X-ray crystal structure analysis reported in this paper have been deposited with the Cambridge Crystallographic Data Center (CCDC) as supplementary publication No. CCDC-160271. Copies of the data can be obtained, free of charge, on application to CCDC, 12 Union Road, Cambridge CB2 1EZ, UK [fax: +44(0)-1223-336033 or e-mail: deposit@ccdc.cam.ac.uk].

1.4.1. (1R,2S)-2-(Dibenzylamino)-1-[ethoxy(oxido)phosphino]-3-phenylpropyl acetate (5a). To a stirred solution of anti-4a (701 mg, 1.5 mmol) in CH₂Cl₂ (4.5 mL) was added Ac₂O (0.50 mL, 4.5 mmol), pyridine (0.38 mL, 4.7 mmol) and DMAP (18 mg, 0.15 mmol) at 0°C and the mixture was stirred for 3 h at room temperature. The mixture was poured into cold water and extracted with CHCl₃. The combined extracts were washed with brine and dried over MgSO₄. Removal of the solvent gave a residue which was chromatographed on silica gel (hexane/ EtOAc=3:1 to 1:1) to give **5a** (650 mg, 93%). This compound was obtained as a mixture of diastereomer in a ratio of 1:1. An oil; 1 H NMR (400 MHz, CD₃OD) δ 7.25– 7.13 (15H, m), 6.86 (0.5H, d with small splits, J=583.7 Hz), 6.70 (0.5H, d with small splits, J=579.8 Hz), 5.53-5.51 (1H, m), 4.08-3.97 (2H, m), 3.76 (1H, d, J=13.7 Hz), 3.75 (1H, d, J=13.7 Hz), 3.72-3.60 (1H, m), 3.56 (1H, d, J=13.7 Hz), 3.52 (1H, d, J=13.7 Hz), 3.25–3.18 (1H, m), 2.99-2.88 (1H, m), 2.03 (1.5H, s), 1.98 (1.5H, s), 1.21 (1.5H, t, J=7.0 Hz), 1.20 (1.5H, t, J=7.1 Hz); ³¹P NMR (162 MHz, CD₃OD) δ 33.12, 32.96; IR (neat) 1748, 1219 cm^{-1} ; EIMS m/z 374 (M⁺-CH₂Ph). High resolution MS calcd for $C_{20}H_{25}NO_4P$ (M⁺ – CH₂Ph): 374.1521. Found: 374.1529.

1.4.2. (1*R*,2*S*)-2-(**Dibenzylamino**)-1-[**ethoxy(oxido**)**phosphino**]-4-methylpentyl acetate (5b). The compound 5b was prepared from 4b (1.56 g, 4.0 mmol) in an analogous manner to that for preparation of 5a. Purification of the residue by column chromatography (hexane/EtOAc=3:1 to 1:1) gave 5b (1.64 g, 95%). This compound was obtained as a mixture of diastereomers in a ratio of 1:1. An oil; ¹H NMR (400 MHz, CD₃OD) δ 7.36–7.25 (10H, m), 7.13 (0.5H, d with small splits, J=577.3 Hz), 7.02 (0.5H, d with small splits, J=569.9 Hz), 5.61–5.59 (1H, m), 4.23–4.09 (2H, m), 3.87 (1H, d, J=6.1 Hz), 3.83 (1H, d, J=6.1 Hz), 3.45–3.37 (3H, m), 2.20 (1.5H, s), 2.19 (1.5H, s), 1.97–1.93 (2H, m), 1.35 (1.5H, t, J=6.9 Hz), 1.32 (1.5H, t, J=7.0 Hz), 0.98 (1.5H, d, J=6.6 Hz), 0.97 (1.5H, d, J=6.6 Hz), 0.57 (1.5H, d, J=6.7 Hz), 0.56 (1.5H, d,

J=6.4 Hz); ³¹P NMR (162 MHz, CD₃OD) δ 33.29, 32.58; IR (neat) 1747, 1219 cm⁻¹; EIMS m/z 432 (MH⁺). High resolution MS calcd for C₂₄H₃₅NO₄P (MH⁺): 432.2303. Found: 432.2299.

1.4.3. (1R,2S)-2-(Dibenzylamino)-1-(dimethoxyphosphoryl)-3-phenylpropyl acetate (6a). A solution of 5a (564 mg, 1.21 mmol), DMSO (0.1 mL, 1.45 mmol) and iodine (30 mg, 0.12 mmol) in THF (4 mL) was stirred for 5 h at 60°C. The mixture was evaporated to give a residue. To a stirred solution of the residue in CH₂Cl₂ (4 mL) was added TMSBr (0.4 mL, 3.03 mmol) and stirred for 12 h at room temperature. After the mixture was concentrated, the residue was dissolved in MeOH (4 mL) and stirred for 2 h at room temperature. Evaporation of the mixture gave a residue. To a stirred solution of the CH₂N₂ in Et₂O (25 mL), prepared from 70 % N-nitroso-N-methylurea (713 mg, 4.84 mmol), was added a solution of the residue in Et₂O/MeOH=10:1 (6.6 mL) at 0°C and the solution was stirred for 30 min at the same temperature. After decomposition of excess CH₂N₂ with AcOH (0.05 mL), the mixture was diluted with H₂O and extracted with Et₂O. The combined extracts were washed with brine and dried over MgSO₄. Removal of the solvent gave a residue which was chromatographed on silica gel (hexane/EtOAc=2:1 to EtOAc) to give **6a** (63 mg, 11%). An oil; $[\alpha]_D^{25} = +7.0$ (c 0.49, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.25–7.01 (15H, m), 5.88 (1H, d with small splits, J=13.2 Hz), 3.84 (2H, d, J=13.8 Hz), 3.75 (3H, d, J=10.8 Hz), 3.65 (3H, d, J=10.6 Hz), 3.56-3.50 (1H, m), 3.31 (2H, d, J=13.8 Hz), 3.22 (1H, dd, J=3.4, 14.7 Hz), 3.01 (1H, dd, J=10.9, 14.7 Hz), 2.18 (3H, s); 13 C NMR (100 MHz, CDCl₃) δ 169.4, 139.7, 138.7, 129.5, 128.7, 128.0, 126.8, 125.9, 65.5 (d, J_{PC} =162.2 Hz), 60.3, 58.0 (d, J_{PC} =5.7 Hz), 53.2 (d, J_{PC} =10.2 Hz), 53.1 (d, J_{PC} =10.7 Hz), 33.4, 21.0, 14.2; ³¹P NMR (162 MHz, CDCl₃) δ 23.56; IR (neat) 1748, 1222 cm⁻¹; EIMS *m/z* 482 (MH⁺). High resolution MS calcd for $C_{27}H_{31}NO_5P$ (M⁺-H): 480.1939. Found: 480.1944.

(1R,2S)-2-(Dibenzylamino)-1-(dimethoxyphosphoryl)-4-methylpentyl acetate (6b). The compound 6b was prepared from **5b** (1.70 g, 4.0 mmol) in an analogous manner to that for preparation of 6a. Purification of the residue by column chromatography (hexane:EtOAc=3:1 to 1:1) gave **6b** (126 mg, 7%). An oil; $[\alpha]_D^{25} = -35.0$ (c 0.85, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.33–7.21 (10H, m), 5.86 (1H, d, J=13.8 Hz), 3.82 (2H, d, J=13.4 Hz), 3.74 (3H, d, J=10.7 Hz), 3.67 (3H, d, J=10.6 Hz), 3.24 (2H, d, J=13.4 Hz), 3.21–3.18 (1H, m), 2.15 (3H, s), 1.99–1.88 (1H, m), 1.81–1.74 (1H, m), 1.45– 1.38 (1H, m), 0.93 (3H, d, J=6.8 Hz), 0.43 (3H, d, J=6.5 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 169.4, 139.4, 129.2, 128.1, 127.0, 126.8, 65.5 (d, J_{PC} =161.5 Hz), 54.0 (d, J_{PC} = 5.9 Hz), 53.3, 53.0 (d, J_{PC} =6.5 Hz), 52.9 (d, J_{PC} =7.2 Hz), 36.3, 24.0, 20.9, 20.6; ³¹P NMR (162 MHz, CDCl₃) δ 23.94; IR (neat) 1748, 1223 cm⁻¹; EIMS m/z 448 (MH^+) . High resolution MS calcd for $C_{24}H_{35}NO_5P$ (MH^+) : 448.2252. Found: 448.2208.

1.4.5. (1*R*,2*S*)-2-(Dibenzylamino)-1-(diethoxyphosphoryl)-3-phenylpropyl acetate (8a). To a stirred solution of 7a (7.81 g, 19.2 mmol) in CH₂Cl₂ (58 mL) was added Ac₂O

(7.5 mL, 57.6 mmol), pyridine (4.6 mL, 59.5 mmol) and DMAP (234 mg, 1.9 mmol) at 0°C and the mixture was stirred for 3 h at room temperature. The mixture was poured into cold water and extracted with Et₂O. The combined extracts were washed with brine and dried over MgSO₄. Removal of the solvent gave a residue which was chromatographed on silica gel (hexane/EtOAc=4:1 to 2:1) to give **8a** (7.31 g, 75%). An oil; $[\alpha]_D^{26}$ = +4.3 (*c* 0.61, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.25–6.99 (15H, m), 5.89 (1H, d with small splits, J=13.3 Hz), 4.14-4.06 (4H, m), 3.99-3.93 (1H, m), 3.86 (2H, d, J=13.9 Hz), 3.57-3.52 (1H, m), 3.30 (2H, d, J=13.9 Hz), 3.00 (1H, dd, J=11.2, 14.7 Hz), 2.20 (3H, s), 1.30 (3H, t, *J*=7.1 Hz), 1.22 (3H, t, J=7.1 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 169.4, 139.8, 138.9, 129.6, 128.7, 128.0, 126.8, 125.9, 65.3 (d, J_{PC} =162.6 Hz), 62.7 (d, J_{PC} =5.3 Hz, 2 carbons), 58.0 (d, J_{PC} =6.0 Hz), 53.1 (2 carbons), 33.5, 21.1, 16.4 (d, J_{PC} =5.7 Hz), 16.3 (d, J_{PC} =5.7 Hz), 15.2; ³¹P NMR (162 MHz, CDCl₃) δ 21.13; IR (neat) 1749, 1222 cm⁻¹; EIMS m/z 510 (MH⁺). High resolution MS calcd for $C_{29}H_{36}NO_5P$ (M⁺): 509.2331. Found: 509.2349.

1.4.6. (1R,2S)-2-(Dibenzylamino)-1-(diethoxyphosphoryl)-4-methylpentyl acetate (8b). The compound 8b was prepared from 7b (1.23 g, 2.8 mmol) in an analogous manner to that for preparation of 8a. Purification of the residue by column chromatography (hexane/EtOAc=3:1 to 1:1) gave **8b** (929 mg, 70%). An oil; $[\alpha]_D^{26} = -37.7$ (c 0.62, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.33–7.20 (10H, m), 5.85 (1H, d, J=13.6 Hz), 4.15–4.06 (4H, m), 4.02-3.94 (1H, m), 3.82 (2H, d, J=13.4 Hz), 3.24 (2H, d, J=13.4 Hz), 2.15 (3H, s), 1.99–1.89 (1H, m), 1.81–1.74 (1H, m), 1.51-1.44 (1H, m), 1.30 (3H, t, J=7.1 Hz), 1.24(3H, t, J=7.1 Hz), 0.93 (3H, d, J=6.8 Hz), 0.42 (3H, d, J=6.8 Hz)J=6.5 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 169.5, 139.6, 129.4, 129.2, 128.1, 127.0, 65.8 (d, J_{PC} =162.3 Hz), 62.6 (2 carbons, d, J_{PC} =6.9 Hz), 54.1 (d, J_{PC} =5.7 Hz), 53.2, 36.3, 24.2, 24.0, 21.0, 20.8, 20.6, 16.5 (d, J_{PC} =5.8 Hz), 16.4 (d, J_{PC} =6.0 Hz); ³¹P NMR (162 MHz, CDCl₃) δ 21.51; IR (neat) 1749, 1224 cm⁻¹; EIMS *m/z* 476 (MH⁺). High resolution MS calcd for $C_{26}H_{38}NO_5P$ (M⁺): 475.2487. Found: 475.2473.

(1R,2S)-2-(Dibenzylamino)-1-(dimethoxyphos-1.4.7. phoryl)-3-phenylpropyl acetate (6a). To a stirred solution of 8a (1.22 g, 2.4 mmol) in CH₂Cl₂ (8 mL) was added TMSBr (1.7 mL, 9.6 mmol) and stirred for 12 h at room temperature. After the mixture was concentrated, the residue was dissolved in MeOH (11 mL) and stirred for 2 h at room temperature. Evaporation of the mixture gave a residue. To a stirred solution of the CH₂N₂ in Et₂O (54 mL), prepared from 70% N-nitroso-N-methylurea (1.40 g, 9.6 mmol), was added a solution of the residue in Et₂O/ MeOH=10:1 (14.9 mL) at 0°C and the solution was stirred for 30 min at the same temperature. After decomposition of excess CH₂N₂ with AcOH (0.1 mL), the mixture was diluted with H₂O and extracted with Et₂O. The combined extracts were washed with brine and dried over MgSO₄. Removal of the solvent gave a residue which was chromatographed on silica gel (hexane/EtOAc=2:1 to EtOAc) to give **6a** (702 mg, 61%). $[\alpha]_D^{25}$ =+7.9 (c 0.96, CHCl₃). The ¹H NMR spectrum was identical with that of the authentic sample prepared from anti-4a.

- **1.4.8.** (1*R*,2*S*)-2-(Dibenzylamino)-1-(dimethoxyphosphoryl)-4-methylpentyl acetate (6b). The compound 6b was prepared from 8b (784 mg, 1.7 mmol) in an analogous manner to that for preparation of 6a. Purification of the residue by column chromatography (hexane/EtOAc=3:1–1:1) gave 6b (406 mg, 55%). $[\alpha]_D^{25}$ =-36.9 (*c* 0.70, CHCl₃). The ¹H NMR spectrum was identical with that of the authentic sample prepared from *anti*-4b.
- 1.4.9. (1R,2S)-2-(Dibenzylamino)-1-hydroxy-3-phenylpropyl(ethyl)phosphinic acid (9). To a stirred solution of anti-3 (187 mg, 0.41 mmol) in CH₂Cl₂ (2 mL) was added TMSBr (0.11 mL, 0.82 mmol) at 0°C and the mixture was stirred for 12 h at room temperature. After the mixture was concentrated, the residue was dissolved in MeOH (1 mL) and stirred for 1 h at room temperature. Evaporation of the solvent gave **9** (161 mg, 93%). Amorphous; $[\alpha]_D^{25} = +23.2$ $(c\ 0.69, MeOH); ^{1}H\ NMR\ (400\ MHz, CD_{3}OD)\ \delta\ 7.61-6.89$ (15H, m), 4.97–4.88 (1H, m), 4.56–4.46 (1H, m), 4.42 (1H, dd, J=5.3, 9.4 Hz), 4.32–4.21 (1H, m), 4.15–4.06 (1H, m), 4.05-3.97 (1H, m), 3.58 (1H, dd, J=4.5, 15.8 Hz), 3.51 (1H, dd, J=7.2, 15.8 Hz), 1.83–1.70 (1H, m), 1.50–1.37 (1H, m), 1.01 (3H, td, J=7.7, 18.2 Hz); ³¹P NMR (162 MHz, CD₃OD) δ 56.02; ¹³C NMR (100 MHz, CD₃OD) δ 138.7, 132.2, 131.7, 131.2, 131.1, 130.6, 130.3, 130.2, 128.6, 65.4 $(d, J_{PC}=108.3 \text{ Hz}), 65.2 (d, J_{PC}=5.5 \text{ Hz}), 56.8, 55.8, 31.8 (d, J_{PC}=5.5 \text{ Hz}), 56.8, 55.8 (d, J_{PC}=5.5 \text{ Hz}), 56.8, 55.8 (d, J_{PC}=5.5 \text{ Hz}), 56.8, 56.8 (d, J_{PC}=5.5 \text{ H$ J_{PC} =7.5 Hz), 19.3 (d, J_{PC} =90.9 Hz), 5.1 (d, J_{PC} =5.8 Hz); IR (KBr) 3208, 1149 cm⁻¹. FABMS m/z 424 (MH⁺). High resolution MS calcd for C₂₅H₃₁NO₃P (MH⁺): 424.2041. Found: 424.2035.

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References

- Patel, D. V.; Rielly-Gauvin, K.; Ryono, D. E.; Free, C. A.; Rogers, W. L.; Smith, S. A.; DeForrest, J. M.; Oehl, R. S.; Petrillo, Jr., E. W. *J. Med. Chem.* 1995, 38, 4557.
- Stowasser, B.; Budt, K.-H.; Jian-Qi, L.; Peyman, A.; Ruppert, D. Tetrahedron Lett. 1992, 33, 6625.
- 3. (a) Arai, T.; Bougauchi, M.; Sasai, H.; Shibasaki, M. *J. Org. Chem.* **1996**, *61*, 2926. (b) Arai, T.; Sasai, H.; Aoe, K.;

- Okamura, K.; Date, T.; Shibasaki, M. *Angew. Chem., Int. Ed. Engl.* **1996**, *35*, 104. (c) Yamada, K.; Arai, T.; Sasai, H.; Shibasaki, M. *J. Org. Chem.* **1998**, *63*, 3666 and references cited therein.
- 4. Yamagishi, T.; Yokomatsu, T.; Suemune, K.; Shibuya, S. *Tetrahedron* **1999**, *55*, 12125.
- Froestl, W.; Mickel, S. J.; Sprecher, G.; Diel, P. J.; Hall, R. G.; Maier, L.; Strub, D.; Melillo, V.; Baumann, P. A.; Bernasconi, R.; Gentsch, C.; Hauser, K.; Jaekel, J.; Karlsson, G.; Klebs, K.; Maître, L.; Marescaux, C.; Pozza, M. F.; Schmutz, M.; Steinmann, M. W.; Riezen, H.; Vassout, A.; Mondadori, C.; Olpe, H.-R.; Waldmeier, P. C.; Bittiger, H. *J. Med. Chem.* 1995, 38, 3313.
- 6. Reetz, M. T. Angew. Chem., Int. Ed. Engl. 1991, 30, 1531.
- 7. Fitch, S. J. J. Am. Chem. Soc. 1964, 86, 61.
- 8. Preliminary communication of this work: Yamagishi, T.; Suemune, K.; Yokomatsu, T.; Shibuya, S. *Tetrahedron Lett.* **2001**, *42*, 5033.
- 31P NMR spectroscopic analysis has been successfully applied to determine the diastereomeric excess of chiral phosphonate derivatives. See: (a) Yokomatsu, T.; Yamagishi, T.; Shibuya, S. *J. Chem. Soc., Perkin Trans. 1* 1997, 1527. (b) Hammerschmidt, F.; Li, Y.-F. *Tetrahedron* 1994, 50, 10253. (c) Kozlowski, J. K.; Rath, N. P.; Spilling, C. D. *Tetrahedron* 1995, 51, 6385.
- 10. In our earlier study, the reaction of 2a with diethyl t-butyl-dimethylsilylphosphite gave anti-β-amino-α-hydroxy-phosphonate with modest selectivity (syn/anti=1:2.8). See: Yokomatsu, T.; Yamagishi, T.; Shibuya, S. Tetrahedron: Asymmetry 1993, 4, 1401.
- 11. The moderate yield of products might be due to partial hydrolysis of the ethyl phosphinate functionality during the work up.
- 12. We confirmed that the ratio of *anti-4a-B* to *anti-4a-A* was not changed after refluxing a solution of *anti-4a* in ethyl acetate for 4 h. Thus, there was no possibility of epimerization from *anti-4a-B* to *anti-4a-A* in the recrystallization.
- Albouy, D.; Brun, A.; Munoz, A.; Etemad-Moghadam, G. J. Org. Chem. 1995, 60, 6656.
- McKenna, C. E.; Higa, M. T.; Cheung, N. H.; McKenna, M.-C. *Tetrahedron Lett.* 1977, 18, 155.
- Yamagishi, T. PhD Thesis, Tokyo University of Pharmacy and Life Science, 1997.
- Altomare, A.; Burla, M. C.; Camalli, M.; Cascarano, G.; Giacovazzo, C.; Guagliardi, A.; Olidori, G. J. Appl. Crystallogr. 1994, 27, 435.